

# DIARECT Newsletter

## Summary

- [New in Autoimmunity: Biotinylated Antigens](#)
- [New Infectious Disease Antigens: \*Bartonella henselae\* and \*Babesia microti\*](#)

Dear Madam or Sir,

As in the past years, DIARECT will attend the largest international trade fair for medical industry, MEDICA, with over 4,900 exhibitors, taking place in Düsseldorf, Germany from November 13 - 16, 2017. More than 127,000 expected visitors will have the opportunity to connect with global clinical market leaders in laboratory medicine and learn about cutting-edge technologies. To read more about the event click [here](#).

DIARECT would like to invite all attendees to stop by our **booth D27 in the temporary hall 3A**.

We are pleased to introduce new additions to our current product lines

- Autoimmunity, including recently launched biotinylated autoantigens
- Infectious Disease Serology
- Allergy

To get an overview of all our products, including the antigens to be presented at this year's MEDICA, have a look at our new product list, available at this link: [product list](#)

Current issue of our Newsletter gives you a first impression and description of the newest additions to our extensive product portfolio.

We are looking forward to seeing you in Düsseldorf.  
DIARECT AG

## New in Autoimmunity: Biotinylated Antigens

Bead-based technologies for *in-vitro* diagnosis provide several advantages compared to traditional single protein measurement techniques (as the gold standard: ELISA). These include high sensitivity, low sample volume, lower costs, all of which allow rapid turnover while generating reproducible data, through ease of performance and elimination of the need for trained personnel and specialized procedures. Multiplex analysis is used to simultaneously look for multiple targets in one sample applying highly standardized assay procedures with automation and high throughput capability.

Increasing awareness of multifactorial nature of various diseases and pathological states calls for simultaneous, time-saving and cost-effective measurement of multiple analytes, also at sample volumes, which are too small for traditional immunoassays. Multiplex systems could provide a cost-effective solution to the omnipresent demand for productivity increases and have the potential to dramatically accelerate biological and biomedical research by enabling the comprehensive analysis of patient samples to become inexpensive, routine and widespread. The rapidly evolving methodologies already widely used in basic research are slowly penetrating the *in-vitro* diagnostic market.

However, limitations of protein microarrays such as inadequate sensitivity, reproducibility, and scalability and especially higher costs of manufacture prevent these approaches from completely replacing standard techniques.

In bead-based arrays (suspension or liquid arrays) usually 5.6  $\mu\text{m}$  polystyrene particles are used, and the protocol is similar to a classic sandwich ELISA. The use of magnetic beads enables the automation of washing steps as well as construction of fully automated analyzers.

As in some second-generation ELISA technologies, bead-based arrays use biotin-conjugation of the antigen and subsequent conjugation to streptavidin-coated surfaces/beads. Using this approach the strength of attachment of antigens to the base is increased, which results in higher assay sensitivities (WHO 2006).

Biotin, formerly designated vitamin H or coenzyme R, is a water-soluble B-vitamin also known as vitamin B7, and is found in small amounts in numerous foods. It acts as an essential coenzyme for four carboxylases that catalyze essential steps in intermediary metabolism (Wallace *et al.* 1999).

Streptavidin is a homo-tetrameric protein (60 KDa) isolated from the actinobacterium *Streptomyces avidinii*. It has been reported to have binding ability similar to the one originally known from avidin, which was originally isolated from egg yolk (Green *et al.* 1990).

Binding of biotin to streptavidin / avidin is one of the strongest non-covalent interactions known in nature, which results from several factors: the formation of multiple hydrogen bonds and van der Waals forces between biotin and the protein, together with surface polypeptide loops that bury the biotin within the protein. Structural alterations at the biotin binding site produce quaternary changes in the streptavidin tetramer, which promote formations in the twisted beta-sheets that link the tetramer subunits (Weber *et al.* 1989).

For validation of DIARECT's biotinylated antigens a bead-based ELISA was performed using magnetic beads (Dynabeads™ M-280 Streptavidin; Thermo Fisher Scientific) coupled with biotinylated antigens. Coupling was performed according to manufacturer's recommendations with suitable modifications made. The standard ELISA protocol using HRP-conjugated secondary antibodies was adapted to the use of magnetic beads which were effectively washed by taking advantage of magnetic separation. Optical density (OD) was determined using a standard plate reader.

Catalog No.	Product name
20100 / 20101	Thyroid Peroxidase (TPO) biotinylated
20200 / 20201	Thyroglobulin (non rec.) biotinylated
20300 / 20301	Ro/SS-A (60 kDa; rec.) biotinylated
20400 / 20401	Ro/SS-A (60 kDa; non rec.; bov.) biotinylated
20500 / 20501	Ro/SS-A (52 kDa) biotinylated
20600 / 20601	La/SS-B biotinylated
20700 / 20701	GBM biotinylated
20800 / 20801	GAD65 biotinylated

#### References:

Green *et al.* (1990) *Methods Enzymol* 184: 51-67  
Wallace *et al.* (1999) *Biochem J* 30: 1-5  
Weber *et al.* (1989) *Science* 243: 85-88  
WHO (2006) *Environmental Health Criteria* 236: 1-333

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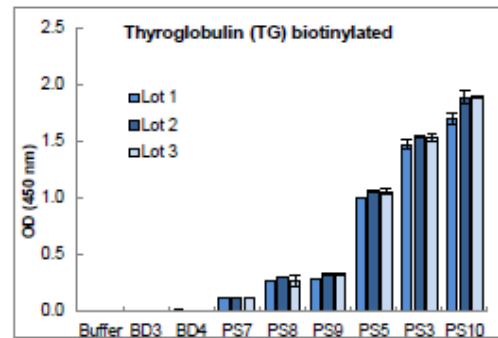
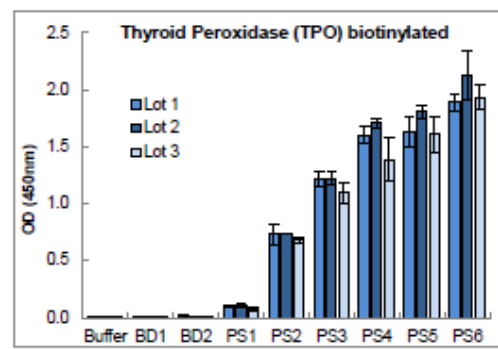


Figure: Three independent lots of TPO (upper graph) and TG (lower graph) were biotinylated and coupled onto streptavidin-coated microspheres. IgG specific responses to the antigens were detected from healthy donor (BD 1 – 4) and patient samples (PS 1 – 10), respectively (double determination). Buffer and non-biotinylated antigens (data available on request) were included as negative controls.

## New Infectious Disease Antigens: *Bartonella henselae* and *Babesia microti*

The rod-shaped, gram-negative and facultative intracellular pathogen *Bartonella henselae* causes several forms of Bartonellosis, including Cat Scratch Disease (CSD) and Bacillary Angiomatosis (BA) (Anderson *et al.* 1995).

CSD was first identified as clinical entity in 1950 in France (Debré *et al.* 1950). A candidate bacterium was observed in 1983 in CSD-patients (Wear *et al.* 1983) and named *Afipia felis* (Brenner *et al.* 1991). In 1992 Regnery *et al.* discovered a new etiologic agent for an infection similar to *A. felis* also identified as causative agent of the clinical syndrome bacillary angiomatosis (Stoler *et al.* 1983). These bacteria in an alpha subdivision of proteobacteria were called *Rochalimaea henselae*, and after later phylogenetic analysis, they were classified under the *Bartonella* genus (Brenner *et al.* 1991).

Due to difficulties in primary culturing of patient samples, indirect immunofluorescence analysis is difficult, which leads to variation in diagnostic test results. The use of recombinant antigens may therefore improve diagnostic accuracy. It is reported that up to 88 or 95% of patients with CSD present antigens against *B. henselae* in sera when tested by an ELISA (Kabeya *et al.* 2003).

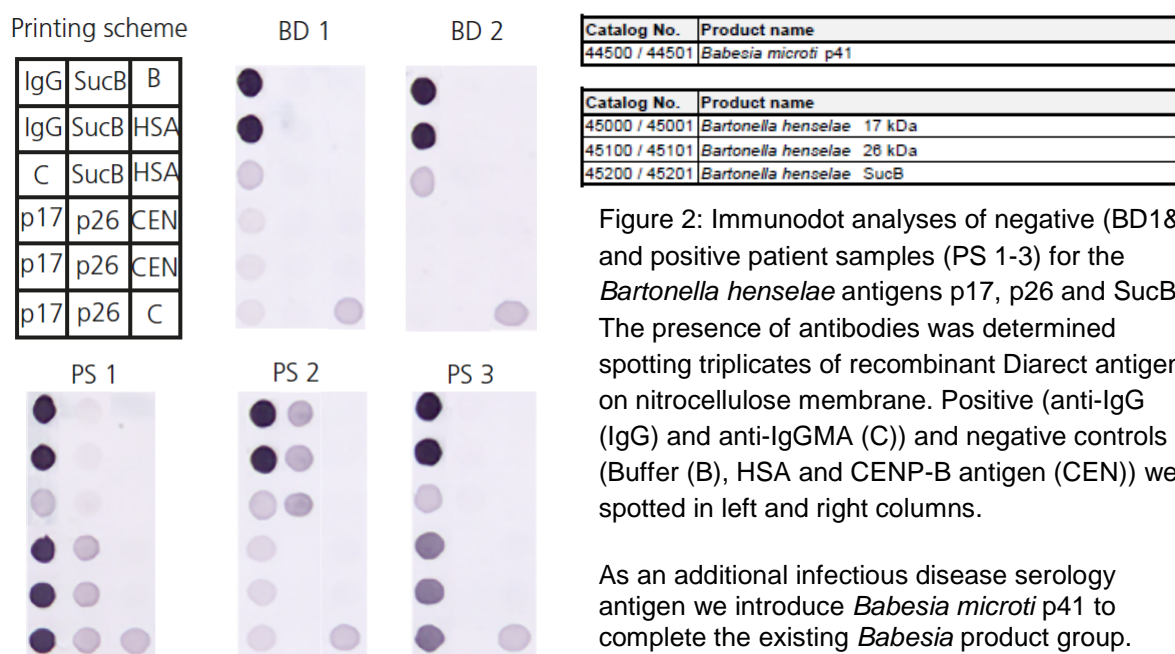
The key factors used as antigens in diagnostics are highly immunoreactive proteins produced in *B. henselae*. Through a Type IV secretory system, *B. henselae* proteins are transported into the host cells, provoking a vascular proliferation in infected endothelial cells (Hoey *et al.* 2009).

The first antigen found to be strongly reactive with sera from CSD patients was p17 (Sweger *et al.* 2000), and it is considered to be a good and species-specific marker for *B. henselae* during early stages of infection. The gene encoding this protein lies in the virB-operon of the Type IV secretion system (Hoey *et al.* 2009). The N-terminus of the protein is hydrophobic, while the rest of the sequence is hydrophilic or neutral and may represent the surface-exposed epitopes. It is presumed to be a bacterial membrane-associated protein containing lysine residues at the N-terminus that interact with phospholipids of the membrane, a typical motif for outer surface proteins. The hydrophobic residues form a membrane-spanning or membrane anchor domain (Anderson *et al.* 1995). As a homolog of the virB5 family, it is strongly suggested to be involved in host recognition by the type IV secretion system,

most likely though protein-protein interactions involving amino acids within the C-terminus (Hoey *et al.* 2009).

Outer membrane protein p26 was identified and characterized by Werner *et al.* (2006). The encoding gene has significant nucleotide identity with orthologs in *Brucellae* spp., *Bartonellae* spp., and several plant-associated bacteria. It is expressed during feline infection, contains a hydrophobic transmembrane region and four dominant antigenic sites (Werner *et al.* 2008).

Molecular characterization of the *Bartonella* spp. gene SucB revealed, that it encodes the immunogenic protein dihydrolipoamide-succinyltransferase, an enzyme of the alpha-ketoglutarate dehydrogenase complex (Gilmore *et al.* 2003; Kabeya *et al.* 2003). Analysis of the SucB protein crystal structure showed that the E2 subunit is composed of 23 subunits arranged with 432 point group symmetry. The catalytic domain catalyzes the transfer of a succinyl group from the S-succinyl dihydrolipoyl moiety to coenzyme A. The protein is divided into an N-terminal domain, an intermediate linker domain, which is in contact with other proteins of the complex, and the enzyme domain in the C-terminus. The active site of the catalytic domain is located in the middle of a channel formed at the interface between two 3-fold related subunits (Knapp *et al.* 1998). Antigenic cross-reactivity of SucB with other *Bartonella* and *Brucella* species is well known, for example the protein was found to be 76.3% identical to the dihydrolipoamide succinyltransferase enzyme (SucB) of *Brucella melitensis* (Litwin *et al.* 2004).



#### References:

- Anderson *et al.* (1995) *J Clinical Microbiol.* 9: 2358–2365  
 Brenner *et al.* (1991) *J Clinical Microbiol.* 29: 2450–2460  
 Debré *et al.* (1950) *Bull Mem Soc Med Hop.* 66: 76–79  
 Gilmore *et al.* (2003) *Infect Immun.* 71: 4818–4822.  
 Hoey *et al.* (2009) *Clin Vaccine Immunol.* 16: 282–284  
 Kabeya *et al.* (2003) *Microbiol and Immunol.* 47: 571–576  
 Knapp *et al.* (1998) *J Mol Biol.* 280: 655–668  
 Litwin *et al.* (2004) *J Med Microbiol.* 53: 1221–1227  
 Regnery *et al.* (1992) *The Lancet.* 340: 557-558  
 Stoler *et al.* (1983) *Am J Clin Pathol.* 80: 714–718  
 Sweger *et al.* (2000) *Clin Diagn Lab Immunol.* 7: 251–257  
 Wear *et al.* (1983) *Science.* 221: 1403-1405  
 Werner *et al.* (2008) *Comp Med.* 58: 375–380  
 Werner *et al.* (2006) *Clin Vaccine Immunol.* 13: 830–836

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## Feedback

If you would like to receive more detailed information on DIARECT products, please follow [this link](#) to our contact and order form.

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